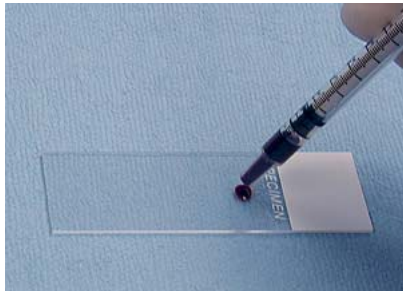


Procedure for Making a Blood Smear

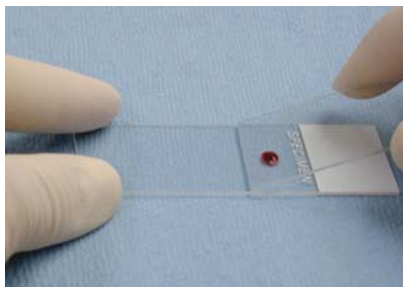
Peripheral blood or potassium EDTA anticoagulated blood (1-2 mg EDTA/1 ml blood) may be used. Smears of peripheral blood must be made immediately. Smears made from EDTA-anticoagulated blood should be made within 2 hours of collection. All specimens must be free of clots.



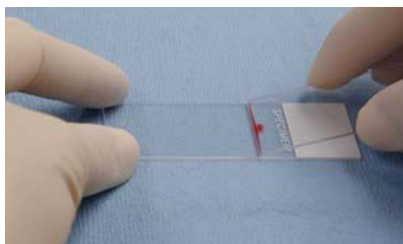
For best results use a Bev-L-Edged slide for both the blood smear and second slide (“spreader”)

Step 1. Using a capillary tube, place a 2-3 mm drop of blood about 1 cm from the frosted end of a clean slide that is on a flat surface.

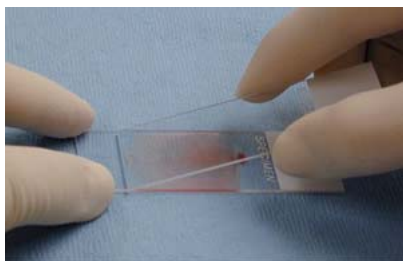
If using a needle and syringe, first remove the needle and then touch the end of the syringe to the slide.



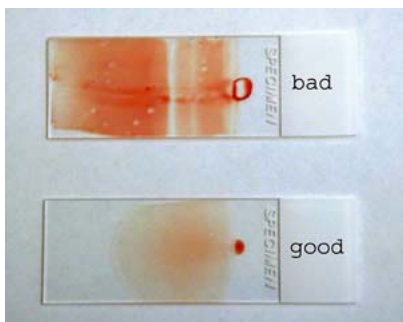
Step 2. With the thumb and forefinger of the right hand, hold the end of the “spreader” against the surface of the first slide at an angle of 30-45 degrees.



Step 3. Draw it back to contact the drop of blood. Allow the blood to spread and fill the angle between the two slides.



Step 4. Push the “spreader slide” at a fast speed forward until all the blood has been spread into a moderately thin film. Apply little to no pressure. Don’t “lift off” spreader slide – push all the way to end of bottom slide.



Note: the film should not cover the entire surface of the slide. In a good film, there is a thick portion and a thin portion and a gradual transition from one to the other. The film should have a smooth, even appearance and be free from ridges, waves, or holes. The end of the smear (the “feathered edge”) should be smooth and even. The edge of the “spreader” must be absolutely smooth. If it is rough, the film has ragged tails containing many leukocytes.